

Paratransgenesis as a tool to block trypanosome transmission by tsetse.

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THIRD FAO/IAEA INTERNATIONAL CONFERENCE ON AREA-WIDE MANAGEMENT OF INSECT PESTS:
INTEGRATING THE STERILE INSECT AND RELATED NUCLEAR AND OTHER TECHNIQUES
VIENNA, 22 - 26 MAY 2017



Outline of the presentation

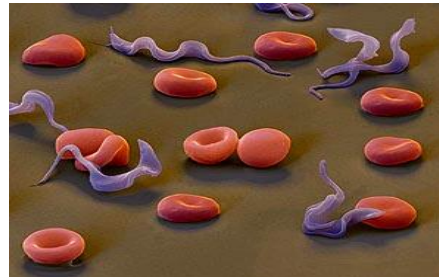
- Background on tsetse-transmitted trypanosomiases and its control
- Potential role of paratransgenic refractory tsetse flies within SIT programs
- Paratransgenesis in tsetse fly: proof-of-concept; current bottlenecks



Tsetse-transmitted African trypanosomiasis

- Parasitic disease(s) caused by *Trypanosoma* sp. (protozoa, kinetoplastidae) in different host species (man, bovine, goat,...).
- Distribution: sub-Saharan Africa

Tsetse fly (*Glossina* sp.)



Trypanosoma sp.

HAT; sleeping sickness

T.brucei gambiense; *T.brucei rhodesiense*



AAT; nagana

T.congolense; *T.vivax*; *T.brucei brucei*



Tsetse-transmitted African trypanosomiasis

■ HAT (in 2015)

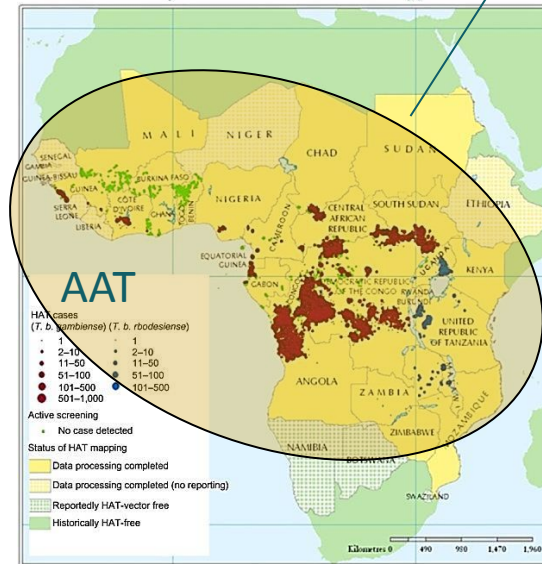
- DRC: 2.351 new cases (>85%);
- CAR: 146;
- 100 cases in surrounding countries
- → remote, rural areas
- > 60 million people are still at risk

WHO: → elimination by 2025?
e.g. TRYP-ELIM program in DRC
(ITM, PNLTHA, LSTM – B&M
funding)

■ AAT

Remains one of the biggest infectious disease constraints to productive livestock rearing in sub-Saharan Africa:

- cattle breeding (increase of mortality and morbidity)
- reduction of meat / milk production: lower income, reduction of nutritional proteins
- losses in animal traction power: reduction of the yields and the surface area that can be cultivated; restricted land usage



Tsetse-transmitted African trypanosomiasis: control

■ HAT:

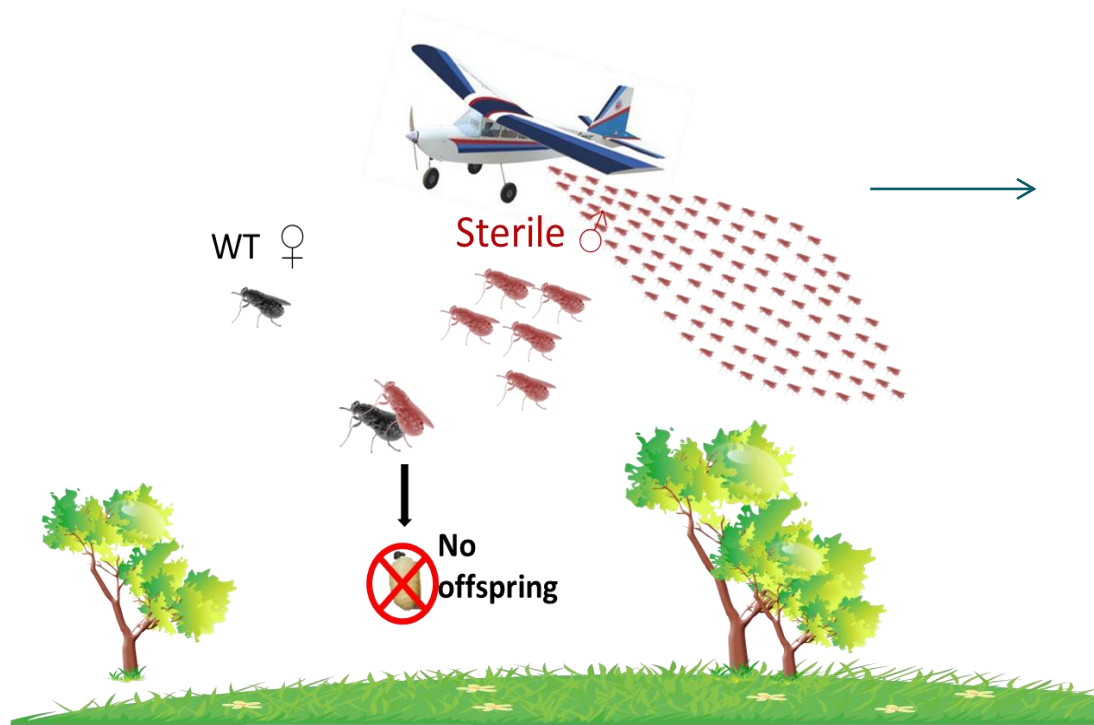
- Active case detection
- Accurate and rapid diagnosis; stage determination
- Treatment: limited amount of drugs available; drug resistance?
- Tsetse fly control: tiny targets



■ AAT:

- Diagnosis by the local vet/farmer,...
- **Treatment:** two main drugs: isometamidium chloride, diminazene aceturate; homidium; important issues: quality of the drugs on the local market; drug resistance.
- Trypanotolerant cattle
- **Tsetse fly control:**
 - use of insecticides: impregnated screens/targets; selective application on animals; sequential aerosol technique – SAT;...
 - **Sterile Insect Technique (SIT)**

Tsetse-transmitted African trypanosomiasis: SIT



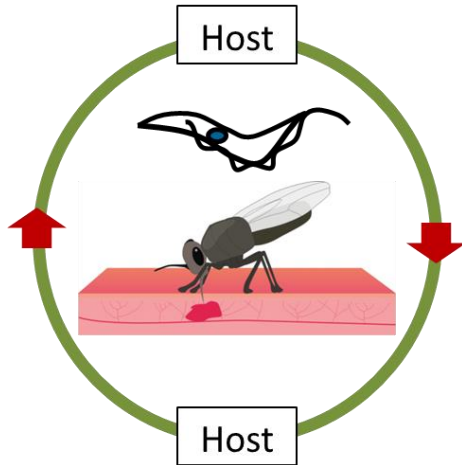
Regular, massive release of potential vectors during a long-time period

↓
Release of 100% trypanosome refractory flies?

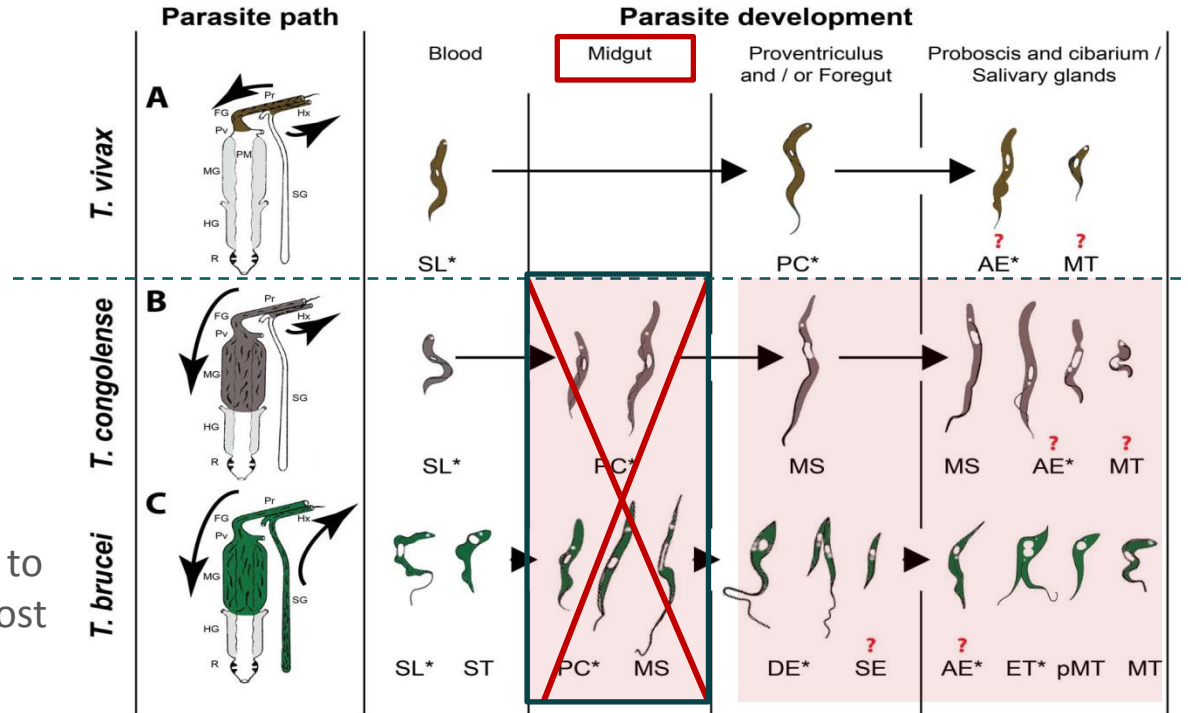
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Paratransgenesis to block trypanosome transmission by the tsetse fly



Trypanosome transmission by the tsetse fly



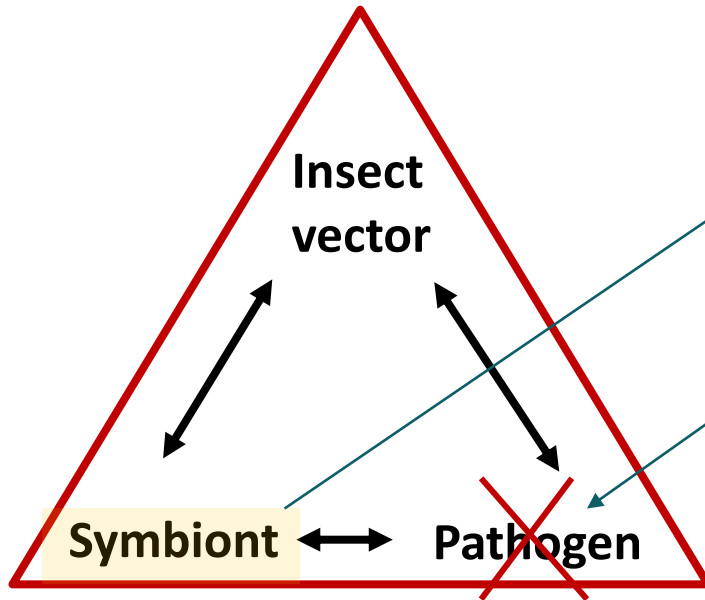
Biological transmission: obligatory developmental cycle in tsetse; simple to complex cfr. *Trypanosoma* species; most of tsetse flies are refractory for trypanosomes



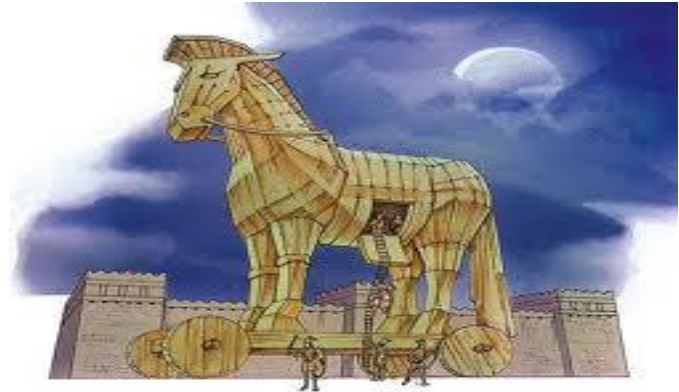
→ Can we achieve this through **paratransgenesis** ?



Paratransgenesis



Trojan horse concept: Genetically modified symbiotic microorganism as an *in situ* delivery system for effector molecules that target the insect-pathogen interplay (paratransgenesis)

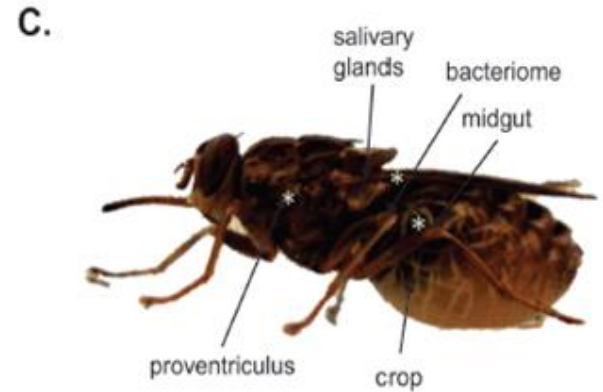
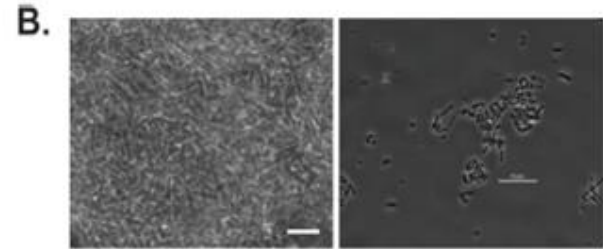
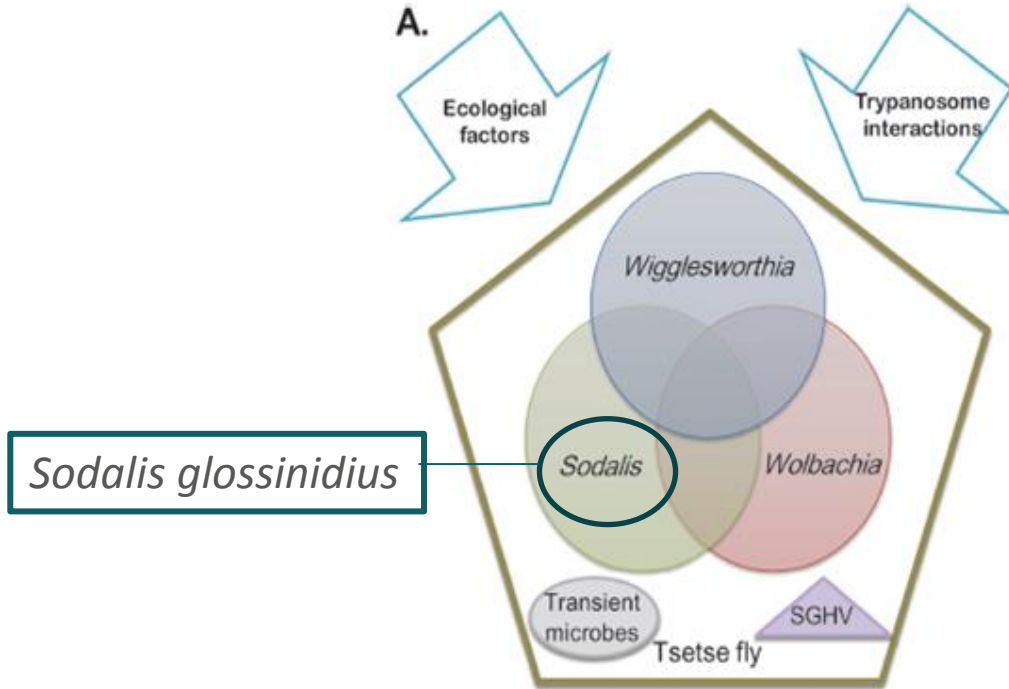


Paratransgenesis

- In order to perform paratransgenesis, there are several requirements:
 - The commensal microorganism (e.g. bacteria) can be grown *in vitro* easily.
 - It can be genetically modified, such as through transformation with a plasmid containing the expression construct/desired gene or by genomic integration.
 - The engineered microorganism is stable and safe.
 - There is a tight and unique association between vector and symbiont
 - Field delivery is possible and easily handled.



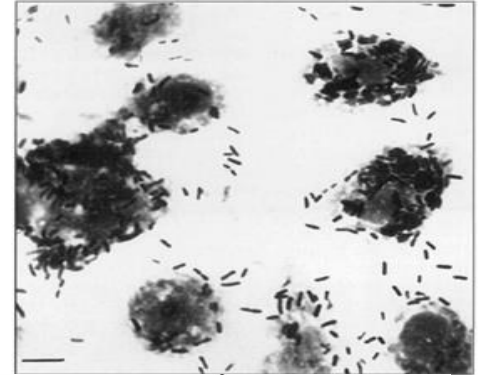
Tsetse candidate microorganisms: microbiome



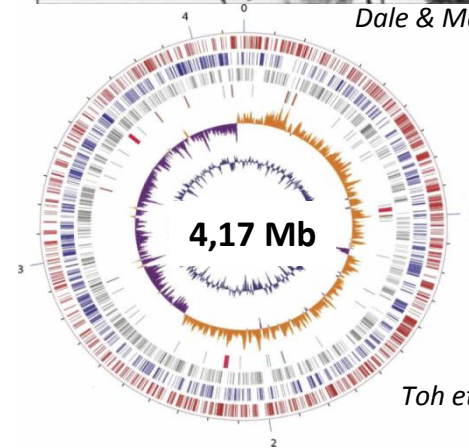
Snyder & Rio, 2013

Facultative commensal: *Sodalis glossinidius*

- Fam. Enterobacteriaceae;
 - Role in tsetse not clear; present in all lab colonies; prevalence in natural populations is highly variable: 0-70%;
 - Vertical transmission to subsequent generations: mother → offspring through the intrauterine nourishment of the larva;
 - Present in the midgut, hemolymph, salivary glands, milk glands; intra- and extracellular;
 - Genome: 4.17Mb; 2,432 protein coding sequences; reduced coding capacity of 51%; 972 pseudogenes;
 - Availability of an *in vitro* culture system and methodologies for genetical modification;
- suitable candidate 'vehicle' for paratransgenesis

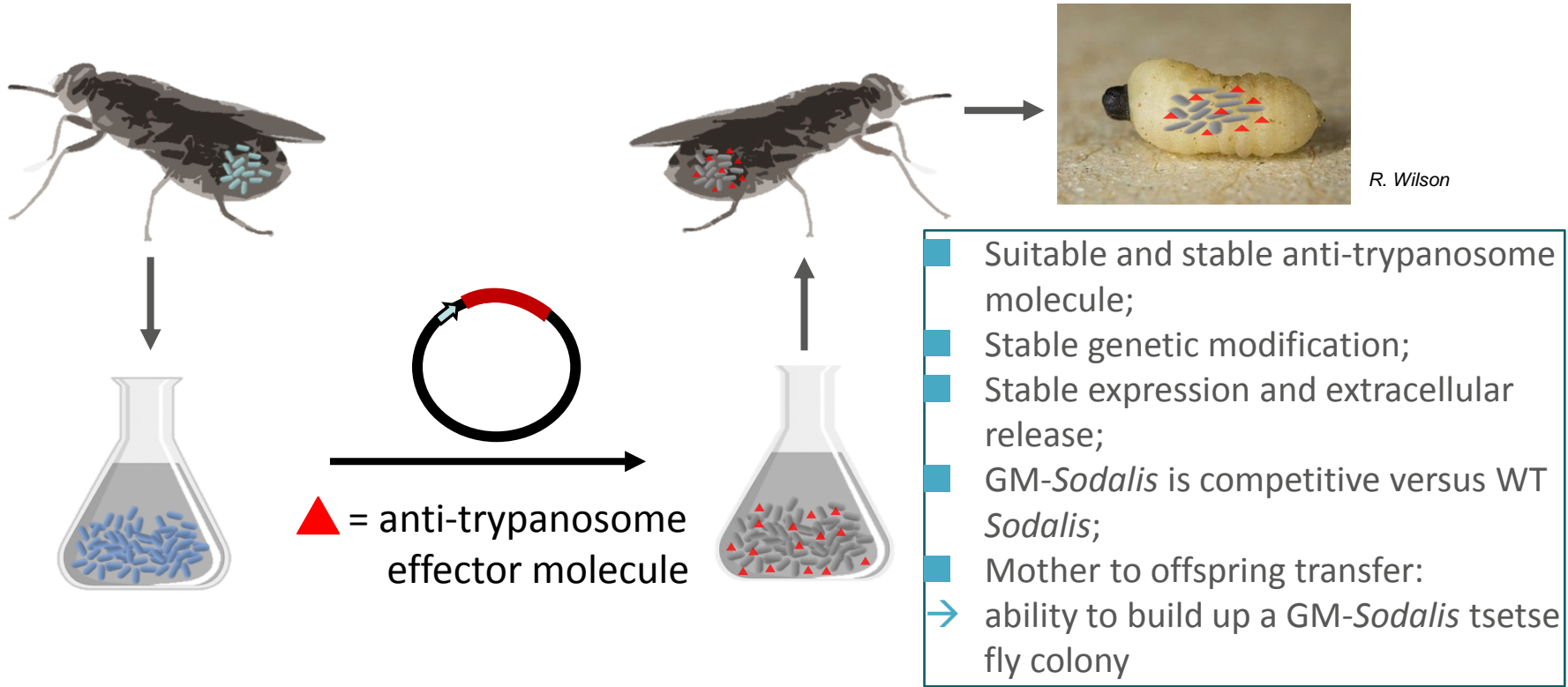


Dale & Maudlin, 1999



Toh et al., 2006

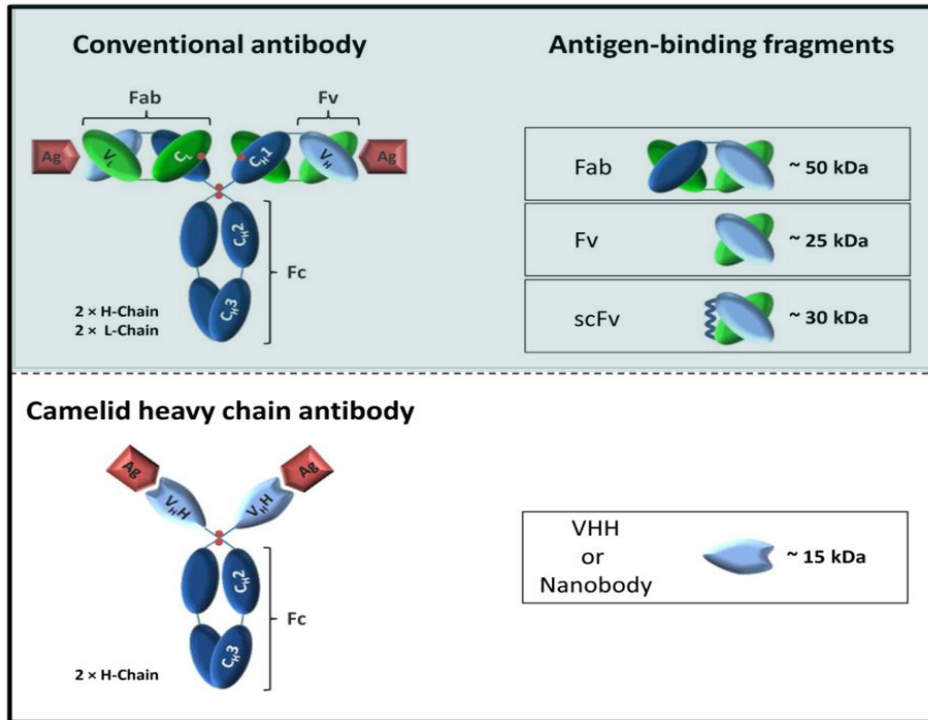
Tsetse – *Sodalis*: paratransgenesis



R. Wilson



Anti-trypanosome molecule: Nanobodies (Nb)



→ Nanobody:

- Single-domain antigen-binding fragments derived from camel heavy-chain antibodies
- Selection through phage display and panning
- Unique epitope repertoire targeting

Advantages:

- Small size (~15 kDa)
- High solubility/stability
- Easy to express; high production yields in *E. coli*
- Nanobodies can be modified: binding affinity; protease resistance,...



Sodalis expression and extracellular release of functional Nb in the tsetse fly

De Vooght et al. *Microbial Cell Factories* 2012, 11:23
<http://www.microbialcellfactories.com/content/11/1/23>



De Vooght et al. *Microbial Cell Factories* 2014, 13:156
<http://www.microbialcellfactories.com/content/13/1/156>

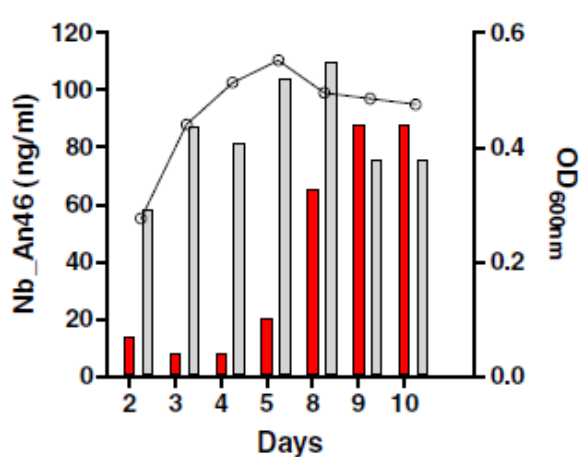


RESEARCH

Open Access

Expression and extracellular release of a functional anti-trypanosome Nanobody[®] in *Sodalis glossinidius*, a bacterial symbiont of the tsetse fly

Linda De Vooght^{1,2}, Guy Caljon^{2,3,4}, Benoit Stijlemans^{3,4}, Patrick De Baetselier^{3,4}, Marc Coosemans¹ and Jan Van Den Abbeele²

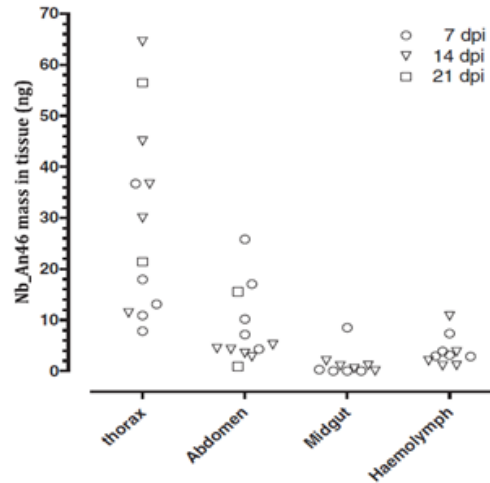


RESEARCH

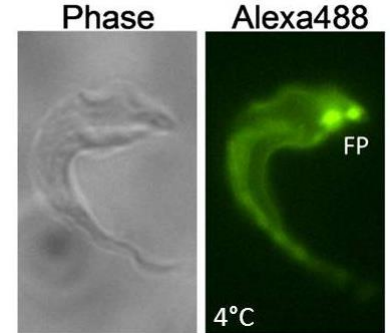
Open Access

Delivery of a functional anti-trypanosome Nanobody in different tsetse fly tissues via a bacterial symbiont, *Sodalis glossinidius*

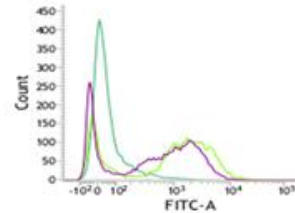
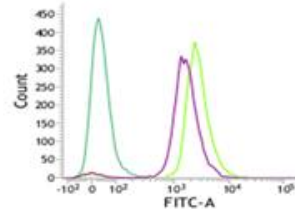
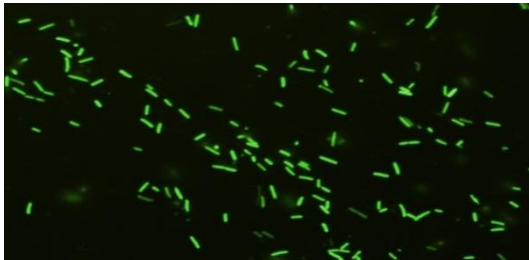
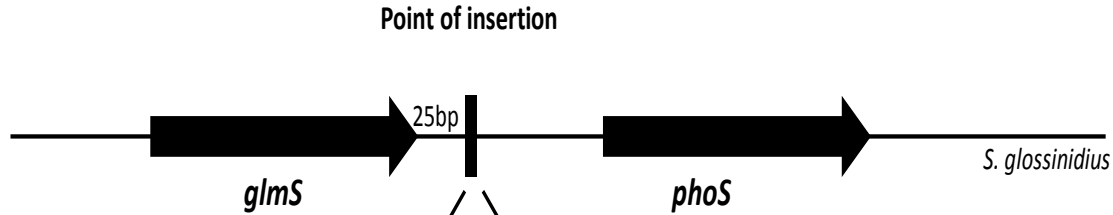
Linda De Vooght^{1*}, Guy Caljon^{1,2,3}, Karin De Ridder¹ and Jan Van Den Abbeele^{1,4*}



Stable expression of functional Nb at the 'ng' level; midgut < level (ELISA problem; midgut proteases)



Stable integration in *Sodalis* genome



Transposon-based system (Tn7) for the site-specific insertion of Nb genes into the chromosome of *Sodalis*:

→ Stable expression of the Nb in the tsetse fly

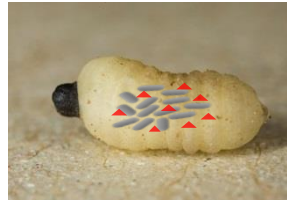
Mother to offspring transfer of GM-*Sodalis*



Needle injection of

fresh-born larvae

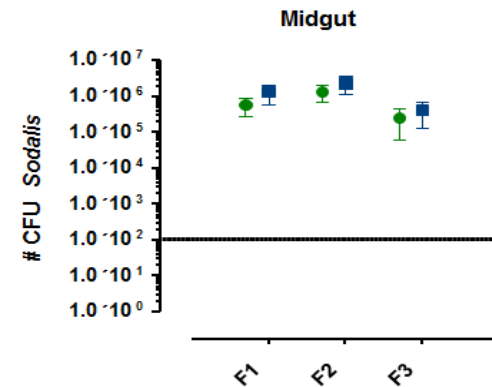
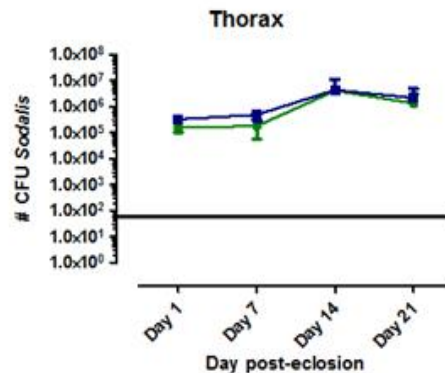
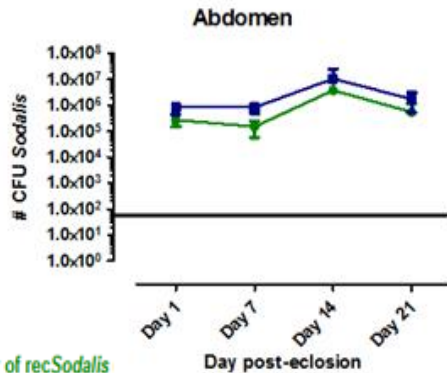
10^6 GM-*Sodalis*



- Intralarval microinjection of GM-*Sodalis* results in **stably colonised adult flies over multiple generations**;
- GM-*Sodalis* is **competitive** vs. WT-*Sodalis*

Adult F₀ female

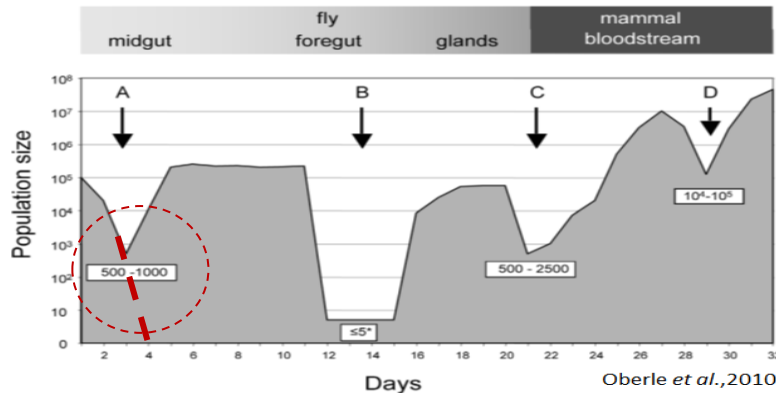
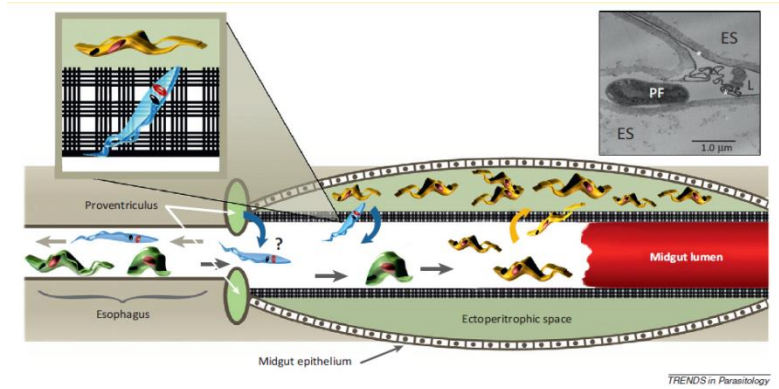
F₁, F₂, F₃ females



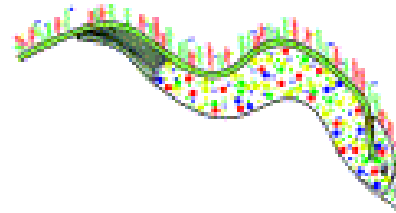
— Total number of rec*Sodalis*

— Total number of *Sodalis*

Blocking trypanosome development in the tsetse midgut?



T. brucei



Pearson, 2001

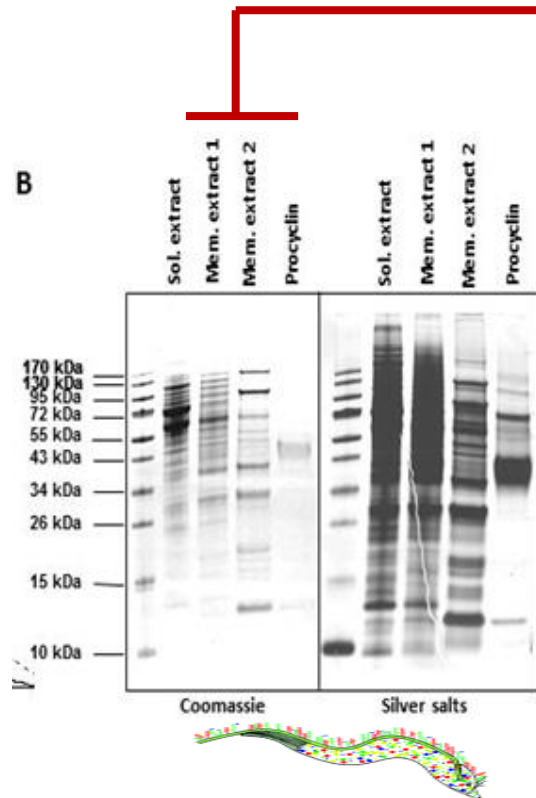
Procyclic stage surface:

- GPEET-procyclin
- EP-procyclin
- Other membrane-bound components?

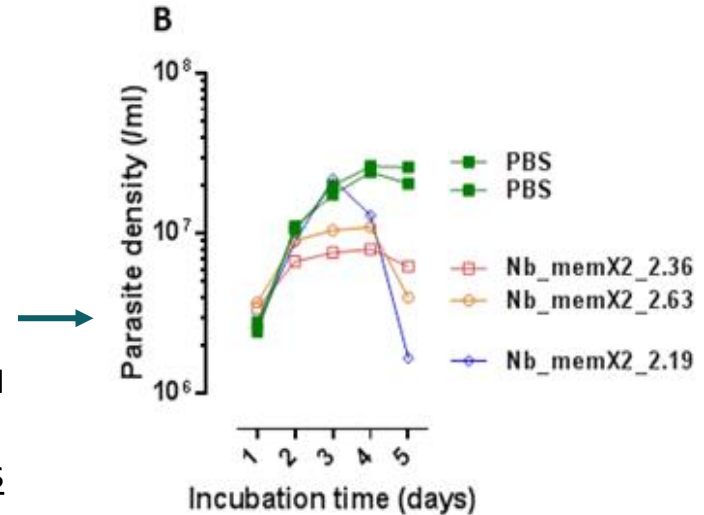
→ Anti-procyclic nanobodies



Selection of anti-procyclic Nbs



Immunisation; Lymphocyte collection; RNA extraction + VHH cDNA; Nb-phage library construction; screening of Nbs for activity on procyclic trypanosomes



Strong trypanolytic effect on *in vitro* cultured procyclics

GM-*Sodalis*/anti-procyclic Nbs in the *T.brucei* infected tsetse

But: *in vivo* in the tsetse fly

<i>Sodalis</i> strain	Infected/total # flies	
	Midgut	Salivary gland
WT <i>Sodalis</i>	17/50	7/17
<i>Sodalis</i> :Nb88	27/53	11/27
<i>Sodalis</i> :Nb63	19/53	9/19
<i>Sodalis</i> :Nb36	21/45	10/21
<i>Sodalis</i> :Nb19	29/43 **	14/29



Enhancement of trypanosome midgut establishment!?



Conclusions

- *Sodalis* is highly suitable for tsetse paratransgenesis;
- Stable genomic integration; expression and release of functional effector molecule in the tsetse fly;
- Efficient transfer to offspring /subsequent generations → possibility to build a tsetse colony with GM-*Sodalis*;
- CURRENT BOTTLENECKs:
 - **highly potent and stable trypanocidal peptide/protein** that blocks trypanosome midgut development?; ideally targeting both *T. brucei* sp. and *T. congolense*.
 - **Impact of irradiation** (sterile males/SIT) on the GM-*Sodalis*?



Acknowledgments



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Nb-work



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Mol.biol work



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tsetse fly rearing



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A. Abdalla, A. Parker, P. Takac



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